









Biomarkers for immuno-oncology: how to maximize chances of therapeutic success

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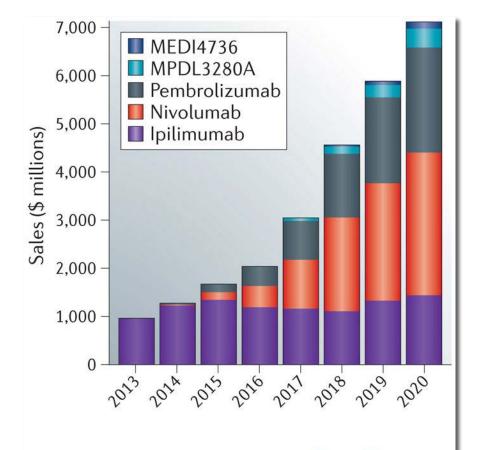
Disclaimer: Any off-label data shown are used to support the educational message of the presentation and not intended to endorse use of any drug in any way

ICIs drive the I-O revolution

Expanding indications

- Melanoma¹⁻³
- Non-small cell lung cancer^{1,2,4,5}
- Renal cell cancer²
- Urothelial (bladder) cancer^{1,2,4}
- Head-neck cancer^{1,2}
- Hodgkin lymphoma^{1,2}
- Merkel cell carcinoma⁶
- All cancers with microsatellite instability (MSI+) regardless of origin⁷

Expanding economical impact⁸



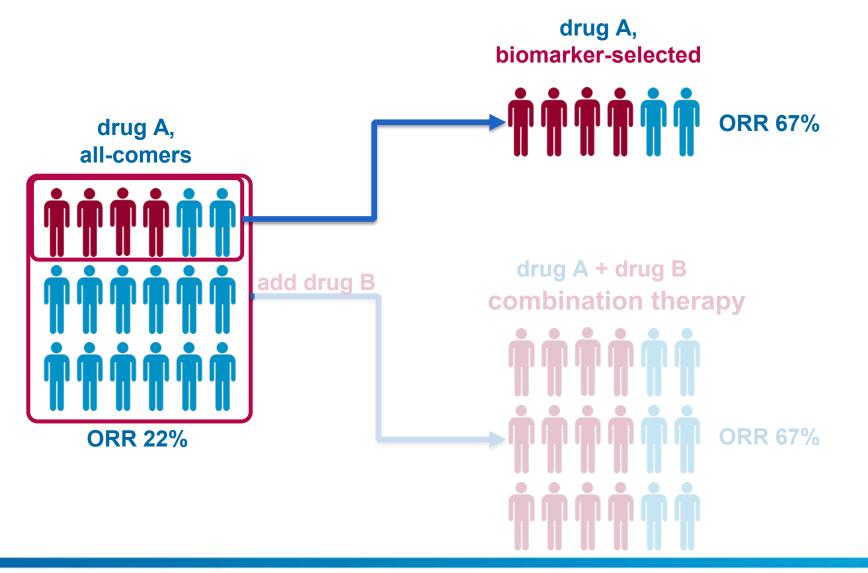
Nature Reviews | Drug Discovery



ICI, immune-checkpoint inhibitors; I-O, immuno-oncology.

1. Keytruda Summary of Product Characteristics. 2. Opdivo Summary of Product Characteristics. 3. Yervoy Summary of Product Characteristics. 5. Imfinzi Summary of Product Characteristics. 6. Bavencio Summary of Product Characteristics. 7. Ciombor KK and Goldberg RM. Drugs 2018;78:155–162. 8. Webster RM. Nature Reviews Drug Discovery 2014;13:883–884.

Why do we need biomarkers in (immuno-)oncology?





Why do we need biomarkers in (immuno-)oncology?

► To increase efficacy of a drug (identify optimal 'responders')

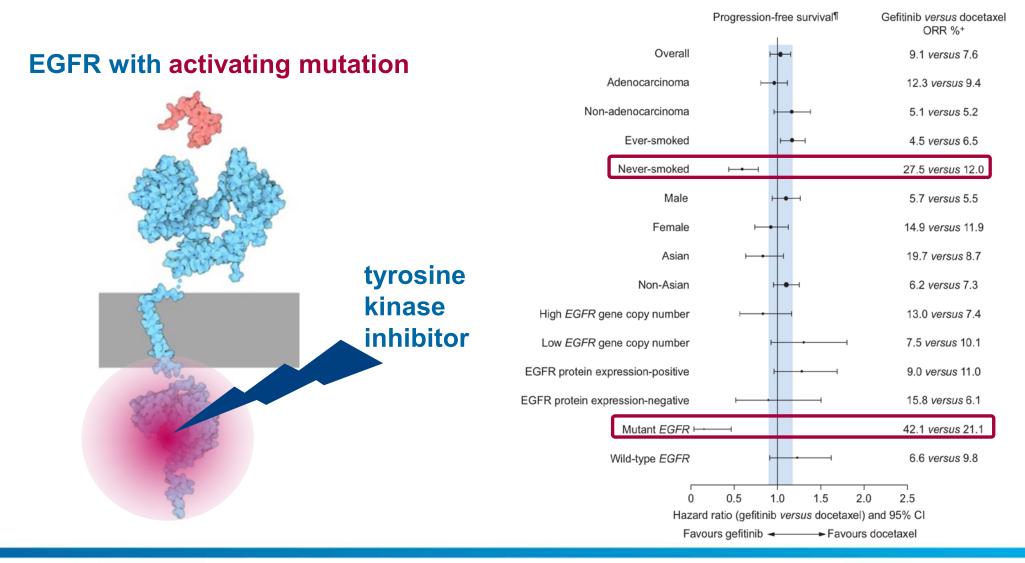
- = avoid unnecessarily treating non-responders

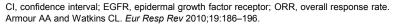
► To avoid toxicity (identify individuals at risk for major side-effects)

To keep healthcare sustainable in the face of constrained government budgets



Going back in time: biomarkers for targeted therapies

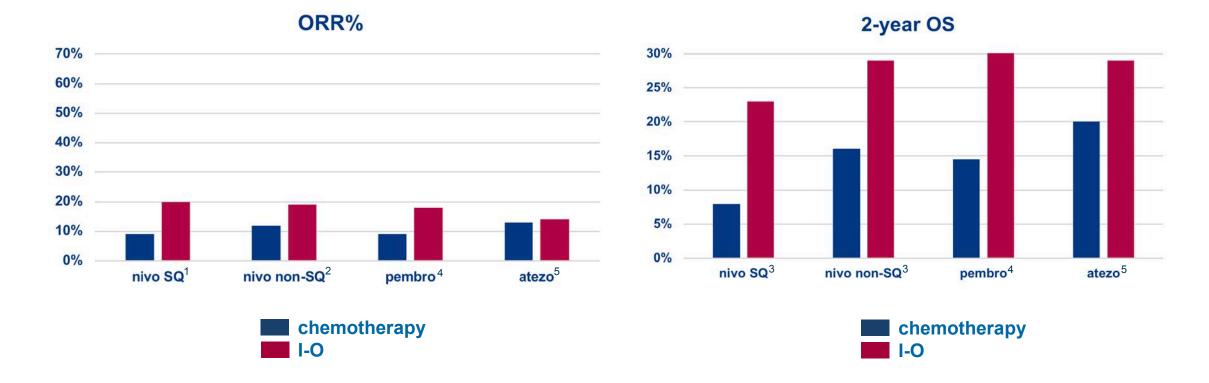






Performance plateau of checkpoints in monotherapy

Immune checkpoint inhibition in monotherapy: performance in second-line NSCLC

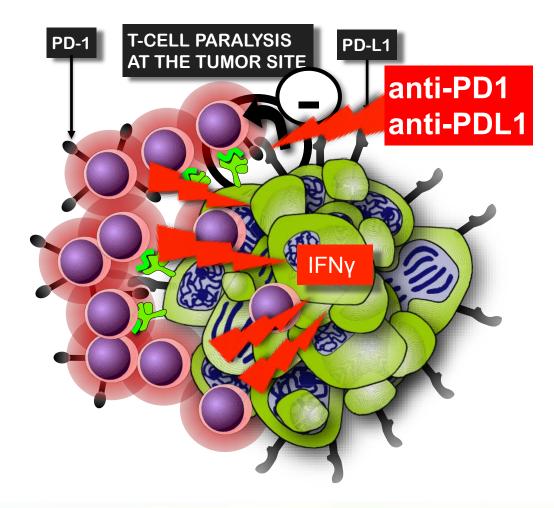




I-O, immuno-oncology; NSCLC, non-small-cell lung carcinoma; ORR, overall response rate; OS, overall survival; SQ, squamous.

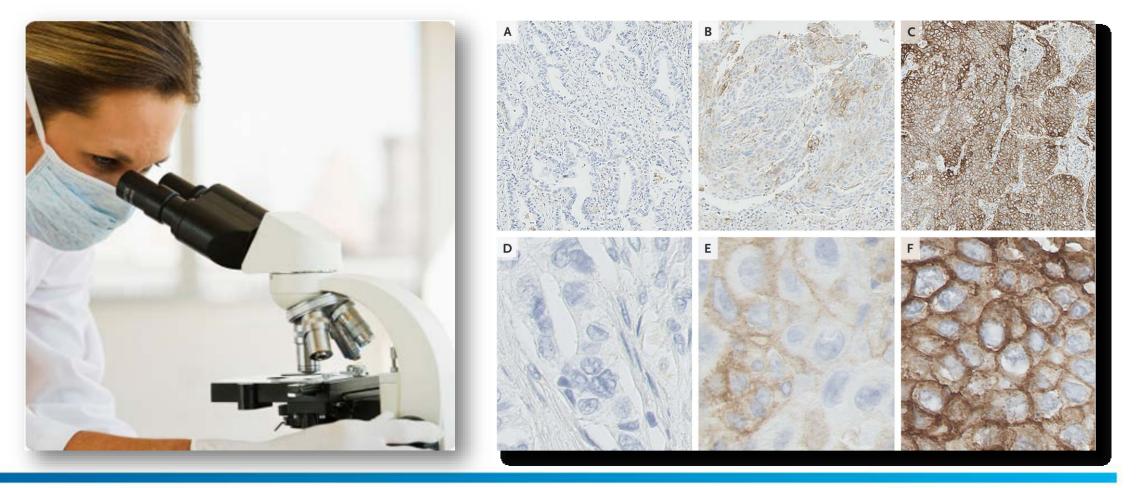
1. Brahmer J et al. N Engl J Med 2015;373:123-35. 2. Borghaei H et al. N Engl J Med 2015;373:1627–1639 3. Horn L et al. J Clin Oncol 2017;35:3924–3933. 4. Herbst R et al. Lancet. 2016;387:1540–50. 5. Rittmeyer A et al. Lancet 2017;389:255–265.

- PD-L1 appears on cancer cells following immune attack
- PD-L1 paralyzes immune cells carrying PD-1





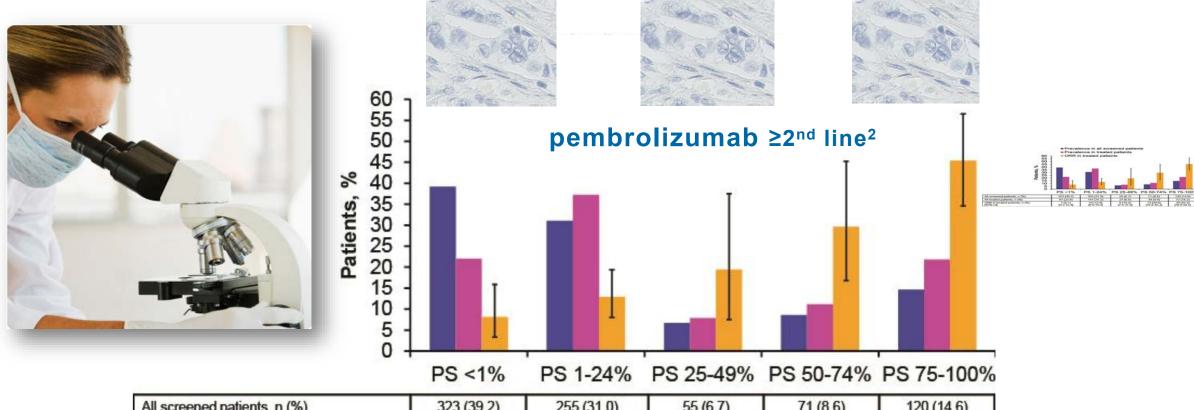
PD-L1 expression is scored on tumor samples (biopsies, some cytological samples)



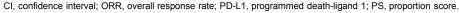
PD-L1, programmed death-ligand 1. Garon EB et al. N Engl J Med. 2015;372:2018–2028.



PD-L1 expression enriches for higher response rate and better survival¹

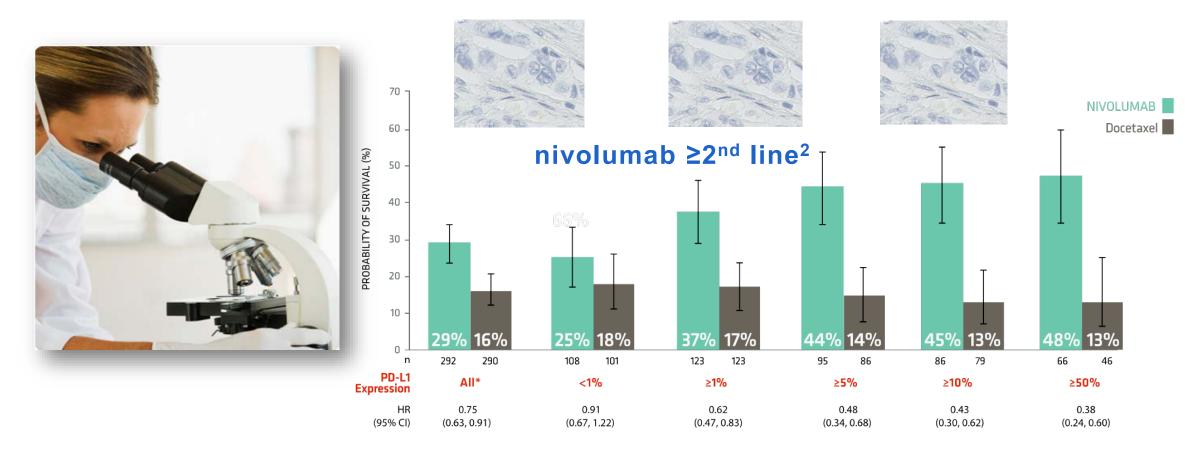


| All screened patients, n (%) | 323 (39.2) | 255 (31.0) | 55 (6.7) | 71 (8.6) | 120 (14.6) |
|--|-----------------------|-------------------------|------------------------|--------------------------|--------------------------|
| All treated patients, n (%) | 87 (22.0) | 147 (37.2) | 27 (6.8) | 39 (9.9) | 72 (18.2) |
| ORR in treated patients, n (%) [95% CI] | 7 (8.1) [3.3-15.9] | 19 (12.9) [8.0-19.4] | 6 (19.4) [7.5-37.5] | 13 (29.6) [16.8-45.2] | 39 (45.4) [34.6-56.5] |



1. Garon EB et al. N Engl J Med. 2015;372:2018–2028. 2. Garon EB et al. N Engl J Med. 2015;372:2018–2028 supplementary material.

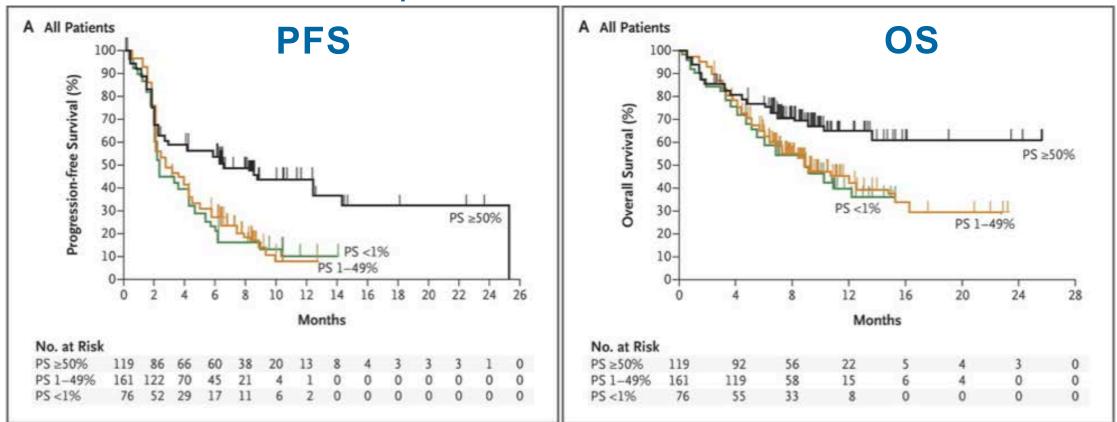
PD-L1 expression enriches for higher response rate and better survival^{1,2}





*All patients include those with no quantifiable PD-L1 expression
**Belgium
CI, confidence interval; HR, hazard ratio; PD-L1, programmed death-ligand 1.
1. Garon EB et al. N Engl J Med. 2015;372:2018–2028. 2. Horn L et al. J Clin Oncol. 2017;35:3924–3933.

PD-L1 expression enriches for higher response rate, better survival

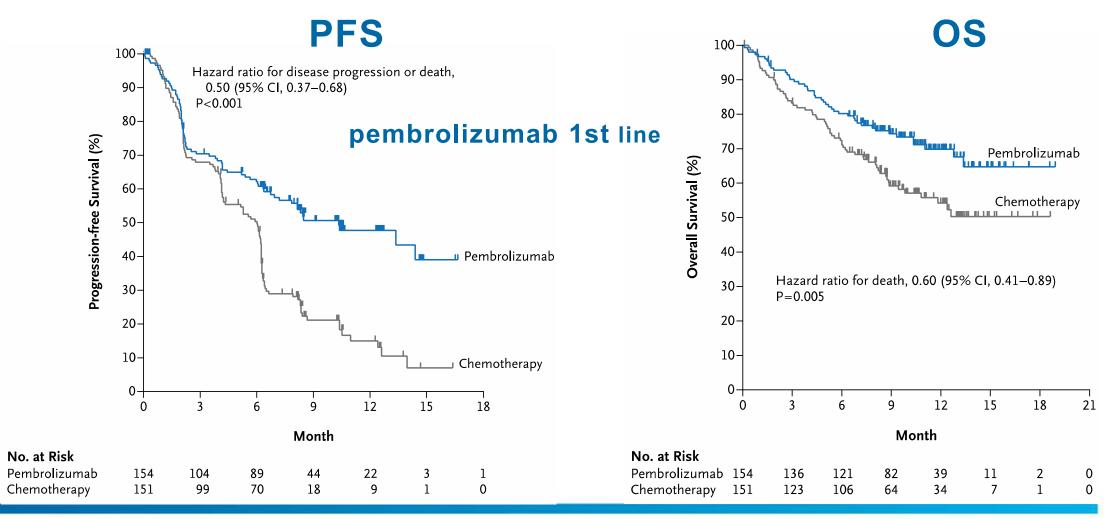


pembrolizumab ≥2nd line

OS, overall survival; PD-L1, programmed death-ligand 1; PFS, progression free survival; PS, proportion score. Garon EB et al. *N Engl J Med.* 2015;372:2018–2028.



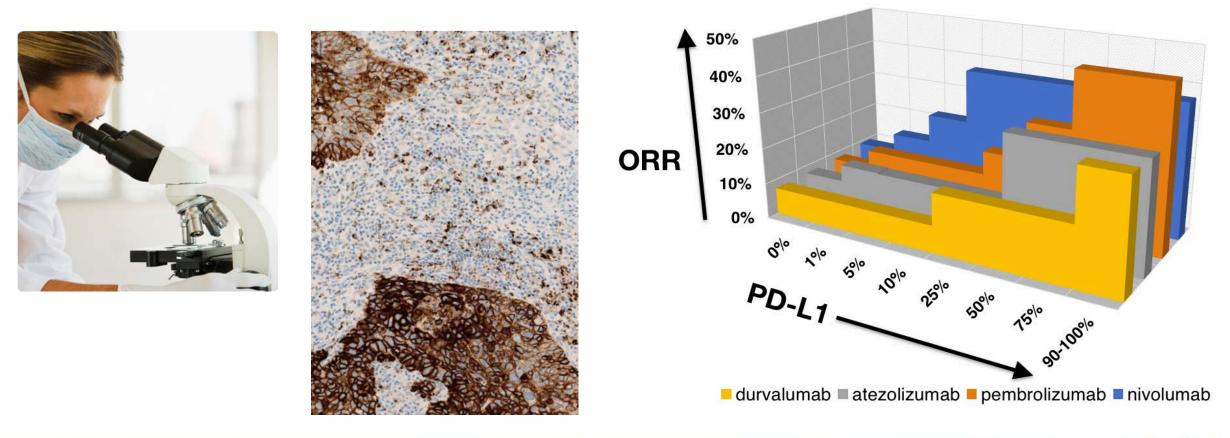
PD-L1 > 50% \rightarrow immunotherapy beats chemotherapy in 1st line



Cl, confidence interval; OS, overall survival; PD-L1, programmed death-ligand 1; PFS, progression free survival Reck M et al. *N Engl J Med.* 2016;375:1823–1833.



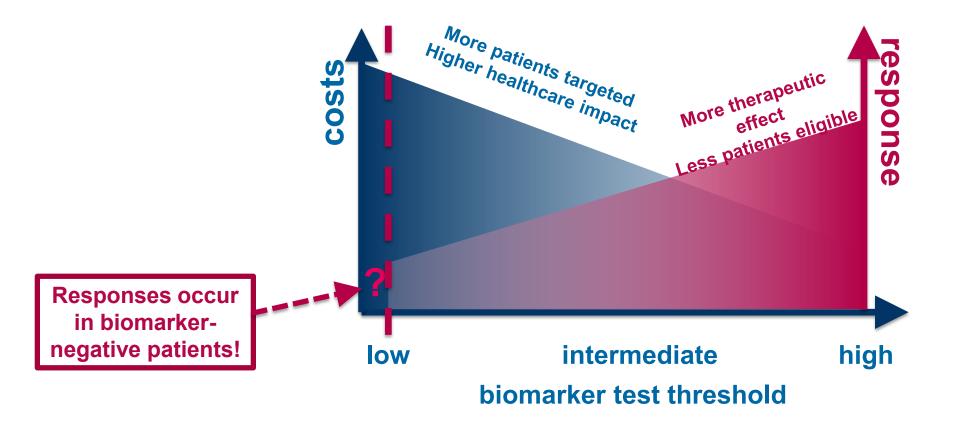
PD-L1 expression enriches for higher response rate, better survival





ORR, overall response rate; PD-L1, programmed death-ligand 1. Vermaelen K et al. *Semin Cancer Biol* 2018;52:166–177.

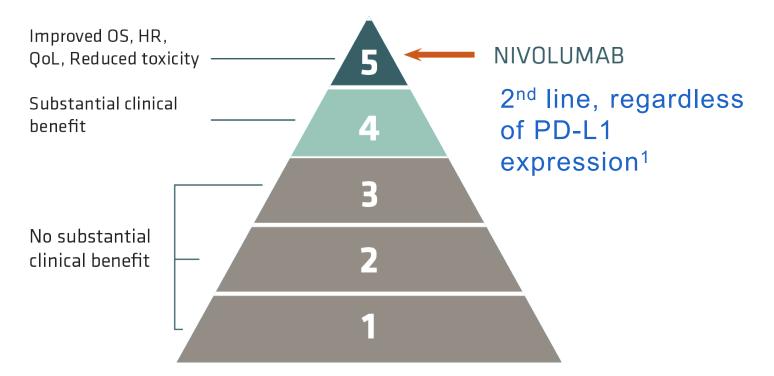
PD-L1 testing does not provide a sharp cut-off to separate responders from non-responders







Clinical benefit can be observed across the whole range of PD-L1 score



ESMO Magnitude of Clinical Benefit Scale (ESMO-MCBS) V1.0^{*2}



*The ESMO Magnitude of Benefit Scale (ESMO-MCBS) is a validated and reproducible tool to assess the magnitude of clinical benefit for cancer medicines. The potential benefits of a new treatment can be summarized as either living longer and/or living better, compared with a control (usually the best current standard care). For second-line treatment of EGFR- and ALK-negative disease, Nivolumab at 3 mg/kg every 2 weeks is recommended in pretreated patients with advanced squamous cell carcinoma (SCC) [I, A; ESMO-MCBS v1.0 score: 5] and it represents a treatment option in pretreated patients with advanced non-squamous cell carcinoma (NSCC) [I, B; ESMO-MCBS v1.0 score: 5]. For NSCC, PD-L1-positive tumour patients benefitted from the use of Nivolumab, compared with docetaxel [II,A] and in PD-L1-negative tumours, Nivolumab and docetaxel showed similar results, with a more favourable toxicity profile for Nivolumab [II,B].

HR, hazard ratio; OS, overall survival; PD-L1, programmed death-ligand; QoL, quality of life. 1. Novello S et al. Ann Oncol. 2016;27(suppl 5):v1-v27. 2. Chery NI et al. Annals of Oncol 2015;26:1547–1573.

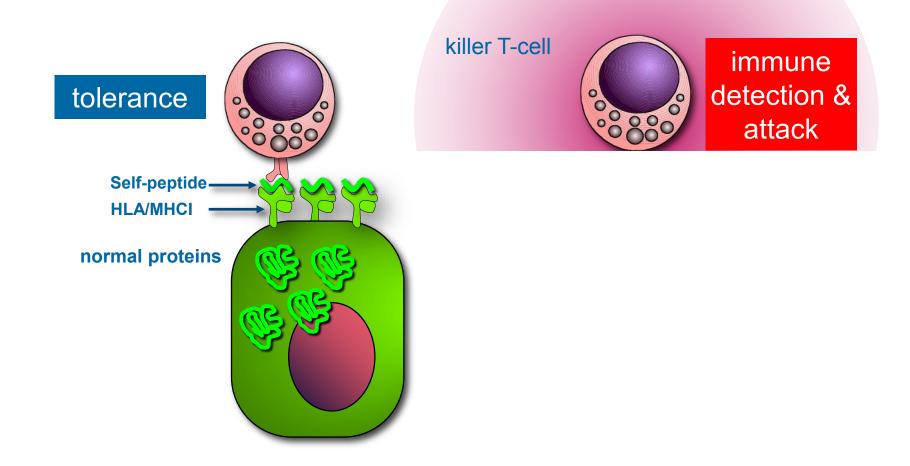
New predictive biomarkers for immuno-oncology

PD-L1: is there anything better on the horizon?



PD-L1, programmed death-ligand 1.

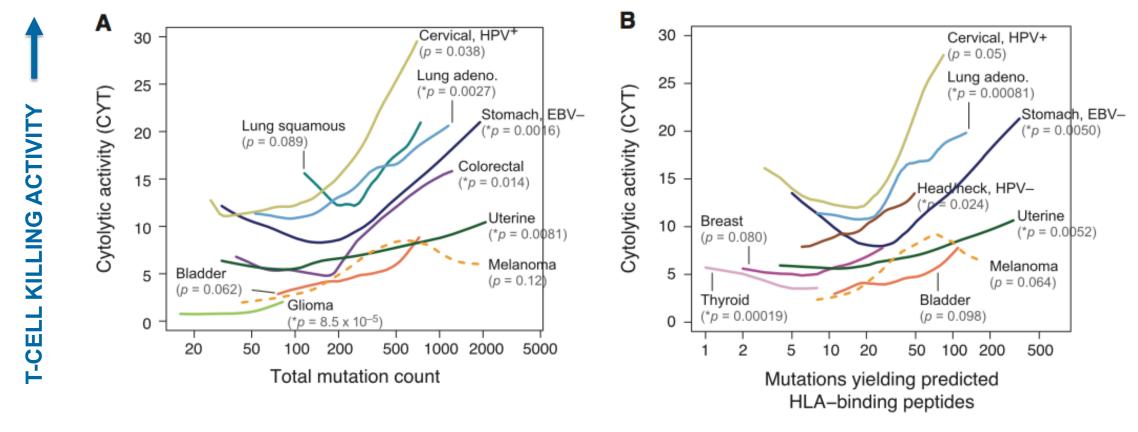
Is response to immunotherapy in the tumor genes?



normal cell



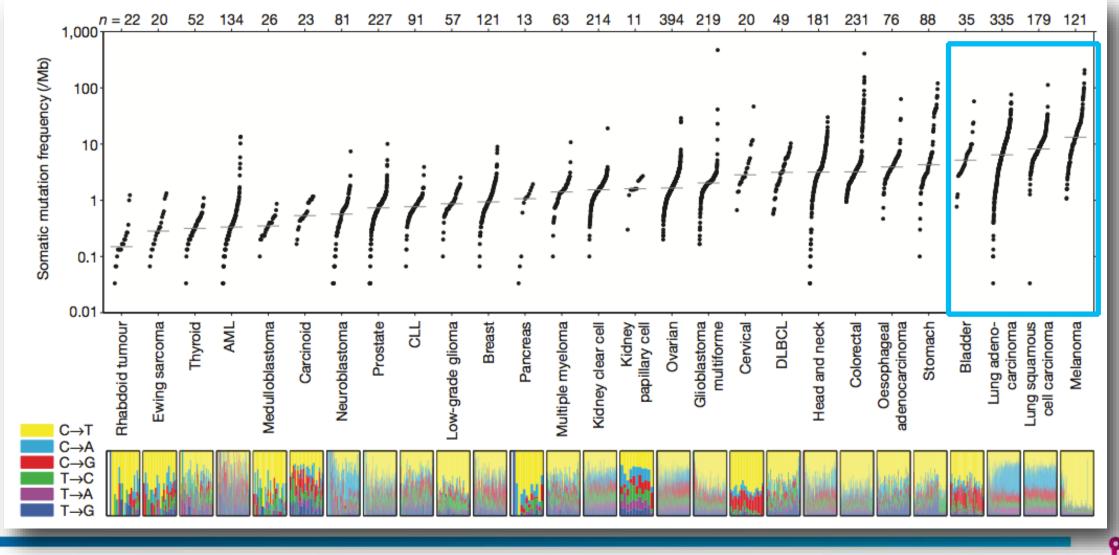
Tumor mutational burden is a driver of immunogenicity







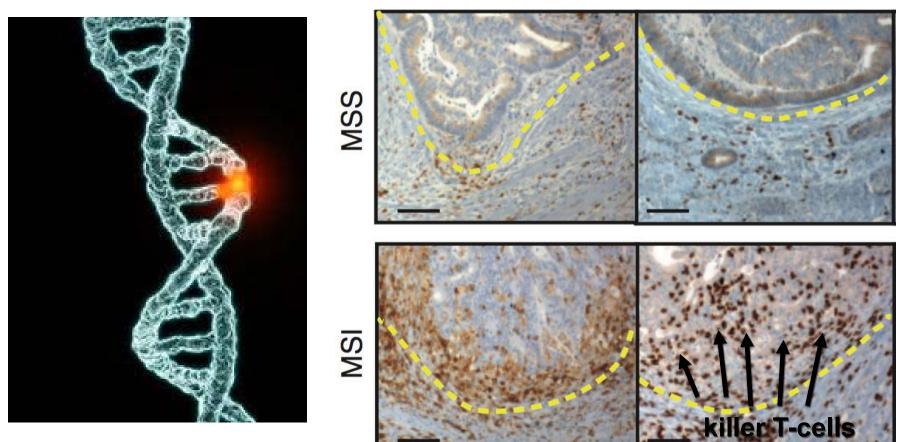
Tumor mutational burden is a driver of immunogenicity

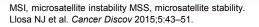


AML, acute myeloid leukemia; CLL, chronic lymphocytic leukaemia; DLBCL, diffuse large B-cell lymphoma Lawrence MS et al. *Nature* 2013;499:214–218.

Tumor mutational burden is a driver of immunogenicity

Cancers with defects in DNA repair (MSI+) are heavily infiltrated with immune cells "MSI-test" as a potential biomarker of response to checkpoint blockade? → investigated

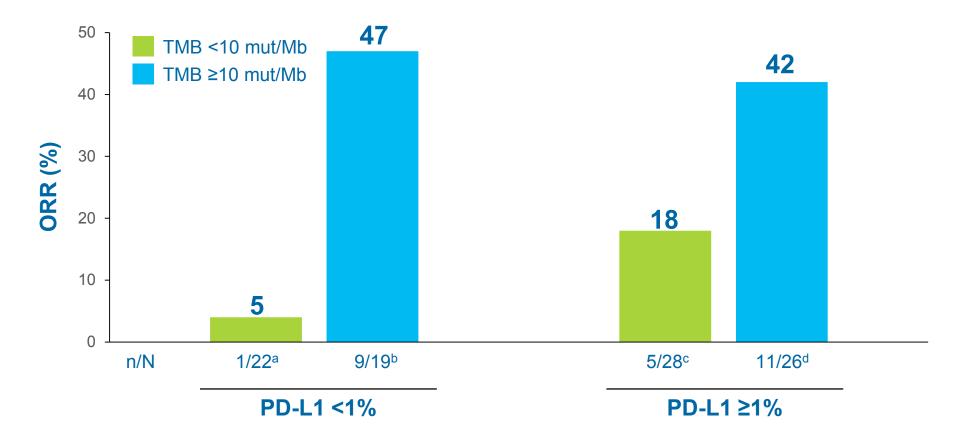






Tumor mutation burden as a predictor of response

Biological & clinical impact of TMB seems to dominate over PD-L1 score





^aCR = 0; ^bCR = 16%; ^cCR = 4%; ^dCR = 4%. CR, complete response; Mb, megabase; Mut, mutation; ORR, overall response rate; PD-L1, programmed death-ligand 1; TMB, tumor mutation burden. Ramalingam S et al. AACR 2018; oral CT078.

Tumor mutation burden as a predictor of response



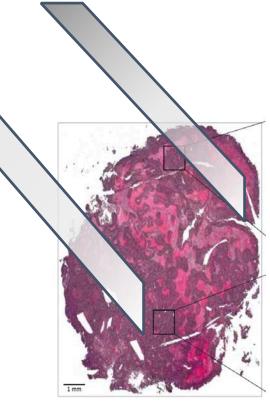
<u>Current</u> limitations of TMB testing for daily clinical practice:

- **Time** to test result +/- 2 weeks
- **Price** per test (e.g. FMI-CDx 3000-4000€)
- Predictive power on **overall survival** warrants further investigation



Limitations of current biomarkers:

- Limited access to tumor tissue in the metastatic setting
- Intratumor heterogeneity





New immunotherapy combinations will require new, specific biomarkers!

Numbers of trials using common combo strategies:

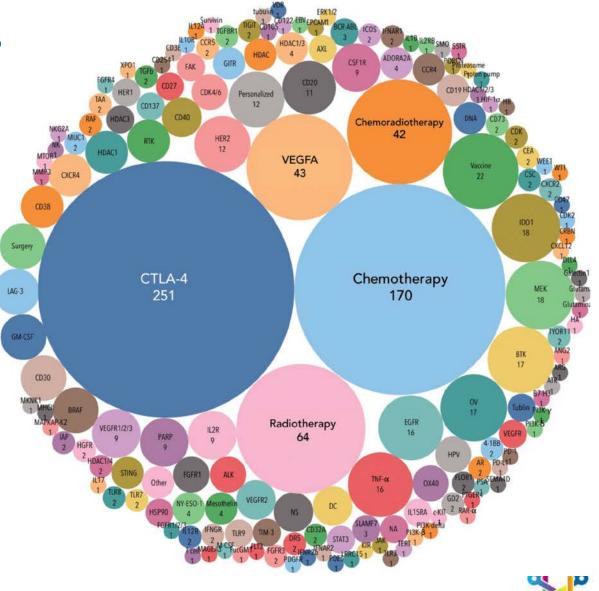
1. Anti-CTLA-4 agents: 251

2. Chemotherapies: 170

3. Radiotherapies: 64

4. Anti-VEGFA agents: 43

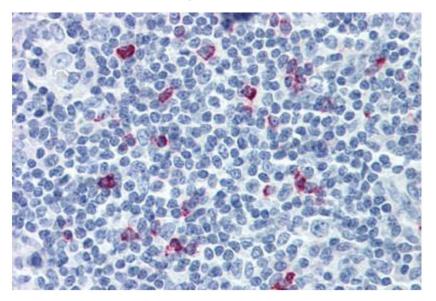
5. Chemoradiotherapy combos: 42



CTLA, cytotoxic T-lymphocyte-associated; I-O, immuno oncology; VEGFA, vascular endothelial growth factor A. Tang J et al. *Annals of Oncol* 2018;29:84–91.

New immunotherapy combinations may require new, specific biomarkers!

Example: LAG3 -new kid on the immune checkpoint block

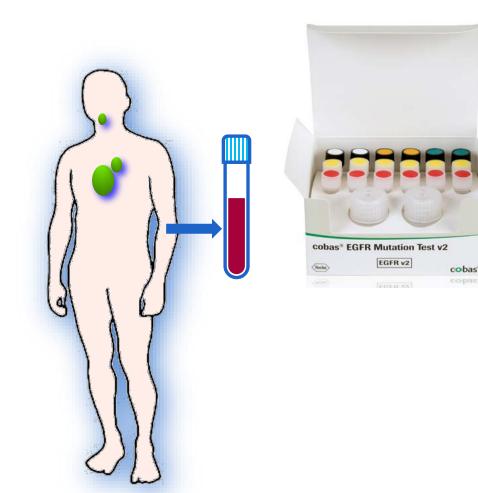


- → Early indications that LAG3-positivity on tumor tissue samples enriches for response to LAG3-targeted combination immunotherapy
- → Under investigation



LAG, lymphocyte-activation gene 3; I-O, immuno oncology. Karim Vermaelen, personal communication, 2018.

Non-invasive biomarkers to predict response to I-O



In contrast to oncogene-targeted therapies, there is no "liquid biopsy" test yet to guide treatment decision in I-O

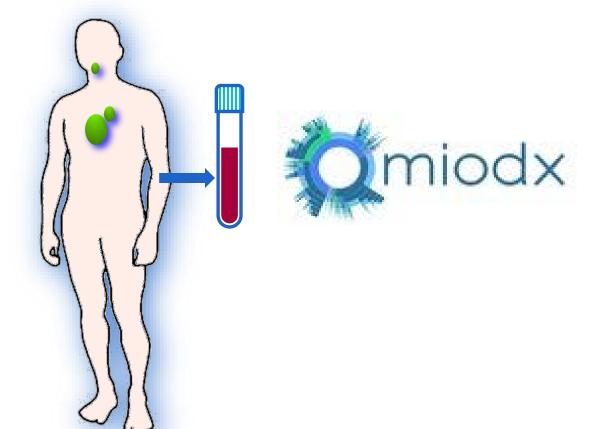
 \rightarrow TMB testing on blood is now in development

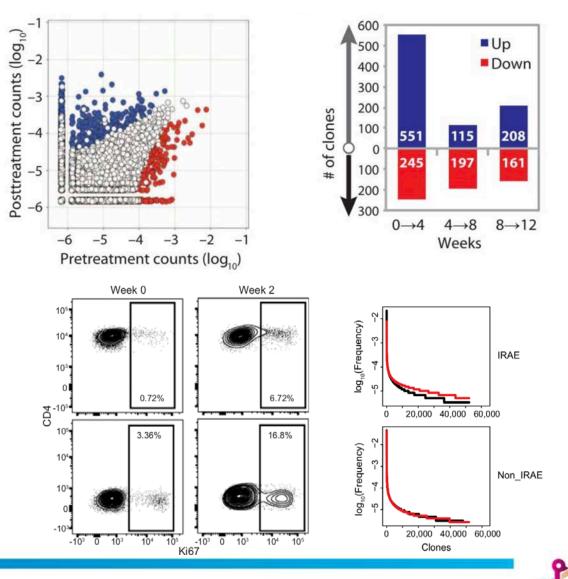


I-O, immuno oncology; TMB, tumor mutational burden. Karim Vermaelen, personal communication, 2018.

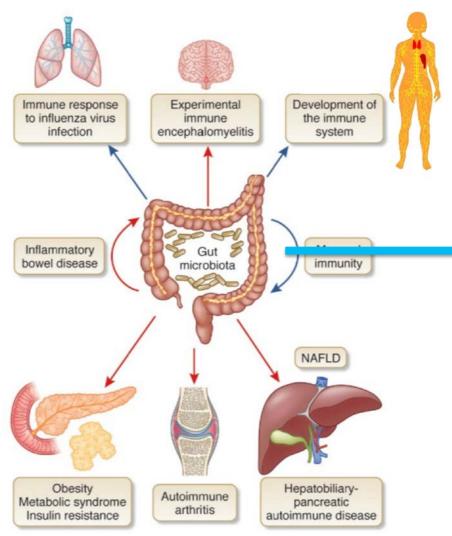
Non-invasive biomarkers to predict response to I-O

Shifts in peripheral blood T-cell clonality can potentially predict response and toxicity to immune checkpoint inhibition









Commensal Bacteria Control Cancer Response to Therapy by Modulating the Tumor Microenvironment

Noriho Iida,¹* Amiran Dzutsev,^{1,2}* C. Andrew Stewart,¹* Loretta Smith,¹ Nicolas Bouladoux,³ Rebecca A. Weingarten,⁴ Daniel A. Molina,⁵ Rosalba Salcedo,¹ Timothy Back,¹ Sarah Cramer,¹ Ren-Ming Dai,^{1,2} Hiu Kiu,¹ Marco Cardone,¹ Shruti Naik,³ Anil K. Patri,⁶ Ena Wang,⁷ Francesco M. Marincola,^{7,8} Karen M. Frank,⁴ Yasmine Belkaid,³ Giorgio Trinchieri,¹†‡ Romina S. Goldszmid¹†‡

SCIENCE VOL 342 22 NOVEMBER 2013



I-O, immuno oncology. Iida N et al. *Science* 2013;342:967–970.



Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota

Marie Vétizou, ^{1,2,3} Jonathan M. Pitt, ^{1,2,3} Romain Daillère, ^{1,2,3} Patricia Lepage, ⁴ Nadine Waldschmitt, ⁵ Caroline Flament, ^{1,2,6} Sylvie Rusakiewicz, ^{1,2,6} Bertrand Routy, ^{1,2,3,6} Maria P. Roberti, ^{1,2,6} Connie P. M. Duong, ^{1,2,6} Vichnou Poirier-Colame, ^{1,2,6} Antoine Roux, ^{1,2,7} Sonia Becharef, ^{1,2,6} Silvia Formenti, ⁸ Encouse Golden, ⁸ Sascha Cording, ⁹ Gerard Eberl, ⁹ Andreas Schlitzer, ¹⁰ Florent Ginhoux, ¹⁰ Sridhar Mani, ¹¹ Takahiro Yamazaki, ^{1,2,6} Nicolas Jacquelot, ^{1,2,3} David P. Enot, ^{1,7,12} Marion Bérard, ¹³ Jérôme Nigou, ^{14,15} Paule Opolon, ¹ Alexander Eggermont, ^{1,2,16} Paul-Louis Woerther, ¹⁷ Elisabeth Chachaty, ¹⁷ Nathalie Chaput, ^{1,18} Caroline Robert, ^{1,16,19} Christina Mateus, ^{1,16} Guido Kroemer, ^{7,12,20,21,22} Didier Raoult, ²³ Ivo Gomperts Boneca, ^{24,25,*} Franck Carbonnel, ^{3,26,*} Mathias Chamaillard, ^{5,*} Laurence Zitvogel^{1,2,3,6} †

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The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients

Vyara Matson,^{1*} Jessica Fessler,^{1*} Riyue Bao,^{2,3*} Tara Chongsuwat,⁴ Yuanyuan Zha,⁴ Maria-Luisa Alegre,⁴ Jason J. Luke,⁴ Thomas F. Gajewski^{1,4}[†]

Matson et al., Science 359, 104-108 (2018) 5 Ja

5 January 2018

Science

REPORTS

Cite as: V. Gopalakrishnan *et al.*, *Science* 10.1126/science.aan4236 (2017).



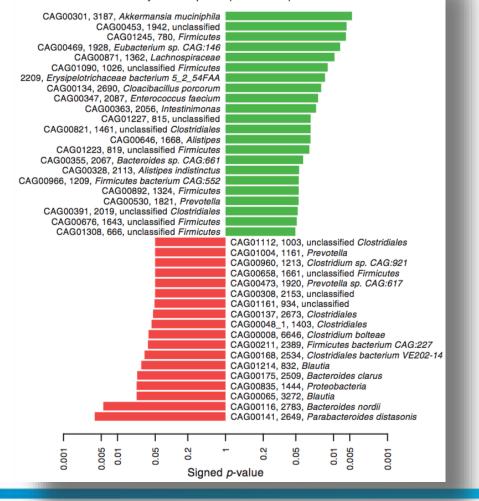
Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients

V. Gopalakrishnan,^{1,2*} C. N. Spencer,^{2,3*} L. Nezi,^{3*} A. Reuben,¹ M. C. Andrews,¹ T. V. Karpinets,³ P. A. Prieto,¹⁺ D. Vicente,¹ K. Hoffman,⁴ S. C. Wei,⁵ A. P. Cogdill,^{1,5} L. Zhao,³ C. W. Hudgens,⁶ D. S. Hutchinson,⁷ T. Manzo,³ M. Petaccia de Macedo,⁶‡ T. Cotechini,⁸ T. Kumar,³ W. S. Chen,⁹ S. M. Reddy,¹⁰ R. Szczepaniak Sloane,¹ J. Galloway-Pena,¹¹ H. Jiang,¹ P. L. Chen,⁹ § E. J. Shpall,¹² K. Rezvani,¹² A. M. Alousi,¹² R. F. Chemaly,¹¹ S. Shelburne,^{3,11} L. M. Vence,⁵ P. C. Okhuysen,¹¹ V. B. Jensen,¹³ A. G. Swennes,⁷ F. McAllister,¹⁴ E. Marcelo Riquelme Sanchez,¹⁴ Y. Zhang,¹⁴ E. Le Chatelier,¹⁵ L. Zitvogel,¹⁶ N. Pons,¹⁵ J. L. Austin-Breneman,¹|| L. E. Haydu,¹ E. M. Burton,¹ J. M. Gardner,¹ E. Sirmans,¹⁷ J. Hu,¹⁸ A. J. Lazar,^{6,9} T. Tsujikawa,⁸ A. Diab,¹⁷ H. Tawbi,¹⁷ I. C. Glitza,¹⁷ W. J. Hwu,¹⁷ S. P. Patel,¹⁷ S. E. Woodman,¹⁷ R. N. Amaria,¹⁷ M. A. Davies,¹⁷ J. E. Gershenwald,¹ P. Hwu,¹⁷ J. E. Lee,¹ J. Zhang,³ L. M. Coussens,⁸ Z. A. Cooper,^{1,3} [P. A. Futreal,³ C. R. Daniel,^{4,2} N. J. Ajami,⁷ J. F. Petrosino,⁷ M. T. Tetzlaff^{6,9} P. Sharma,^{5,19} J. P. Allison,⁵ R. R. Jenq,³ # J. A. Wargo,^{1,3}#**</sup>



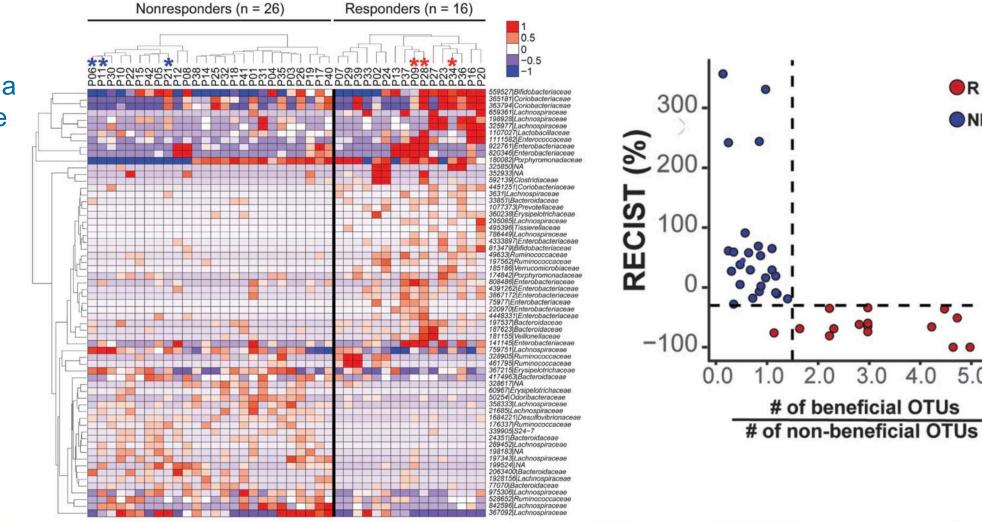
Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors

Bertrand Routy,^{1,2,3} Emmanuelle Le Chatelier,⁴ Lisa Derosa,^{1,2,3} Connie P. M. Duong,^{1,2,5} Maryam Tidjani Alou,^{1,2,3} Romain Daillère,^{1,2,3} Aurélie Fluckiger,^{1,2,5} Meriem Messaoudene,^{1,2} Conrad Rauber,^{1,2,3} Maria P. Roberti,^{1,2,5} Marine Fidelle,^{1,3,5} Caroline Flament,^{1,2,5} Vichnou Poirier-Colame,^{1,2,5} Paule Opolon,⁶ Christophe Klein,⁷ Kristina Iribarren,^{8,9,10,11,12} Laura Mondragón,^{8,9,10,11,12} Nicolas Jacquelot,^{1,2,3} Bo Qu,^{1,2,3} Gladys Ferrere,^{1,2,3} Céline Clémenson,^{1,13} Laura Mezquita,^{1,14} Jordi Remon Masip,^{1,14} Charles Naltet,¹⁵ Solenn Brosseau,¹⁵ Coureche Kaderbhai,¹⁶ Corentin Richard,¹⁶ Hira Rizvi,¹⁷ Florence Levenez,⁴ Nathalie Galleron,⁴ Benoit Quinquis,⁴ Nicolas Pons,⁴ Bernhard Ryffel,¹⁸ Véronique Minard-Colin,^{1,19} Patrick Gonin,^{1,20} Jean-Charles Soria,^{1,14} Eric Deutsch,^{1,13} Yohann Loriot,^{1,3,14} François Ghiringhelli,¹⁶ Gérard Zalcman,¹⁵ François Goldwasser,^{9,21,22} Bernard Escudier,^{1,14,23} Matthew D. Hellmann,^{24,25} Alexander Eggermont,^{1,2,14} Didier Raoult,²⁶ Laurence Albiges,^{1,3,14} Guido Kroemer,^{8,9,10,11,12,27,28*} Laurence Zitvogel^{1,2,3,5*} Enriched in R: Objective response (PR and SD)
 Enriched in NR: Objective response (PD or death)





Could microbiome profiling provide a useful predictive biomarker?





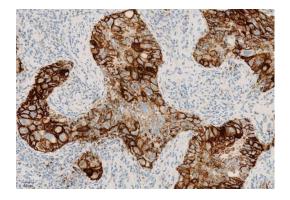
OR

NR

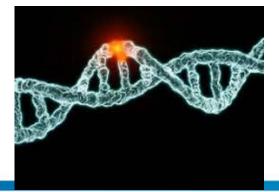
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PD-L1, other immune markers

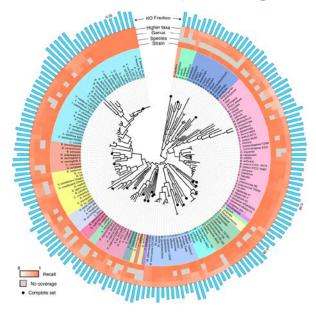


Total mutational burden





Microbiome profiling?





Karim Vermaelen, personal communication, 2018.

Biomarkers in I-O: conclusions / take home messages

Biomarkers should allow us to deliver "precision immuno-oncology"

Identify responders / exclude non-responders (economical burden!)

Challenges for the future:

- Can we design the ideal predictive test: cost-effective, non-invasive, accurate, simple to interpret?
- Can we also find biomarkers to predict severe toxicity?
- ► Will every combination therapy require its own predictive test??
- ► Our understanding of cancer immunology keeps growing → leads to ever more complex candidate biomarkers (e.g. gut microbiota)



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